Version 1.2, Nov 2002

Reviewer/Date		Tige	Conn	ing ha	m/ (6-18-	10
Sr. Review/Da	te	Chi	15 (1 can	di	6/25/	10
Lab Report #				874-		1 1	
Project #	61	071	000	16-	12		

.0	Laboratory Deliverable Requirements	
	1.1 Laboratory Information : Was all of the following provided in the laboratory report? Check items received.	
	Name of Laboratory W Address P Project ID Phone #	Sample identification – Field and Laboratory
	☐ Name of Laboratory ☐ Address ☐ Project ID ☐ Phone # Client Information: ☐ Name ☐ Address ☐ Client Con	ntact (IDs must be cross-referenced)
ACTIO	ON: If no, contact lab for submission of missing or illegible information.	
	1.2 Laboratory Report Certification Statement	Yes [No [] N/A [] Comments:
Does ti	ne laboratory report include a completed Analytical Report Certification in the required	format?
ACTIO	N: If no, contact lab for submission of missing certification or certification with correct	format.
	1.3 Laboratory Case Narrative:	Yes [No [] N/A [] Comments:
B	Narrative serves as an exception report for the project and method QA/QC performance.	Narrative includes an explanation of each discrepancy on the Certification Statement.
ACTIO	N: If no, contact lab for submission of missing or illegible information.	
	1.4 Chain of Custody (COC) copy present of completed COC?	Yes No No N/A Comments:
Does th	e laboratory report include a copy of the completed Chain of Custody forms containing all samples in this SDG?	
NOTE:	Olin receives and maintains the <i>original</i> COC.	
ACTIC	N: If no, contact lab for submission of missing completed COC.	

1.5 Sample Receipt Information (Cooler Receipt Form): Were each of the following tasks completed and recorded upon receipt of the sample(s) into the laboratory? Sample temperature confirmed: must be 1° – 10° C. (If samples were sent by courier and delivered on the same day as collection, temperature recontainer type noted Condition observed PH verified (where applicable) Field and lab IDs cross referenced
Container type noted Condition observed pH verified (where applicable) Field and lab IDs cross referenced
CTION: If no, contact lab for submission of missing or incomplete documentation.
1.5.1 Were the correct bottles and preservatives used? Yes No NA Common NA
Water - 1 Liter amber bottle/cool to 4°C Soil - 8 oz soil jar/cool to 4°C
ACTION: If no, inform senior chemist. Document justification for change in container/volume (if applicable), qualify positive and non-detect data (J) if cooler temperature exceeds 10°C. Rejection of data requires professional judgment.
1.5.2 Were all samples delivered to the laboratory without breakage? Yes No N/A Common N/A Common N/A N/A No N/A N/A No No No No No No No N
1.5.3 Does the Cooler Receipt Form or Lab Narrative indicate other problems with sample receipt, condition of the samples, analytical problems or special Yes No NA Common circumstances affecting the quality of the data?
1.6 Sample Results Section: Was the following information supplied in the laboratory Yes [No [N/A [Compreport for each sample?
Field ID and Lab ID Clean-up method //A Matrix Date and time collected Preparation method Date of preparation/extraction/digestion of Units (soils must be reported in dry weight)
CTION: If no, contact lab for submission of missing or incomplete information.

Me Me	1.7 QA/ for each thod blank	QC Information: Was the sample batch? results LCS recoveries	following information provided in the laboratory report as MS/MSD recoveries and RPDs Surrogate Not Subalties		No []	N/A [_]	Comments:
ACTIO	ON: If no,	contact lab for submission of	of missing or incomplete information.				***
2.0	Holding	Times		Yes []	No [N/A [_]	Comments:
	Have an	y technical holding times,	determined from date of collection to date of analysis, b	een exceede	d?		
		ater samples, the holding to raction and 40 days from ex	ime is 7 days from sampling to extraction and 40 days straction to analysis.	from extrac	tion to ana	lysis. For soi	I samples, the holding time is 14 days from
ACTI detect	ON: If tee results. Fo	chnical holding times are ex r soil samples professional j	ceeded, qualify all positive results (J) and non-detects (U) udgement will be used to determine if rejection is necess	JJ). For wate sary.	r samples th	nat are grossly	exceeded (>2X hold time) reject (R) all non
3.0	Laborato	ory Method		Yes	No [_]	N/A [_]	Comments:
	3.1	Was the correct laborate	ory method used?				
		Water Extraction Soil Extraction Semi-volatile Organics	3510C or 3520C 3540C or 3550B 8270C				
		no, contact project manage requested method.	ger to inform Client of change; request variance fr	om Client;	contact lal	poratory to p	rovide justification for method change
	ΓE: The (Are the practical quanti SOW □ QAPP QAPP and MADEP QA/QC 'erify proper PQLs were use	tation limits the same as those specified by the Lab? C Guidelines provides PQLs for semi-volatile organic d for each data set.	Yes 🔟	No [_]	N/A [_]	Comments:
ACTI	ON: If no,	evaluate change with respec	ct to sample matrix, preparation, dilution, moisture, etc. l	f sample PQ	L is indetern	ninate, contac	t lab for explanation.
							<u> </u>

3.3	3	Are the appropriate parameter results present for each sample in the SDG?	Yes [1	No [_]	N/A [_]	Comments:
ACTION:	If no,	check Request for Analysis to verify if method was ordered and COC to verify that it	was sent, an	d contact la	b for resubm	ission of the missing
3.	4	Were Tentatively Identified Compounds (TICs) reported?	Yes []	No [N/A [_]	Comments:
are requ guidance reported	iired. e, Til ' as T	re only required for samples with full MADEP target list. Determine if TICs MADEP requires that all TICs be reported to the LCS. Per the MADEP Cs, which are identified as aliphatic hydrocarbons, do not have to be TICs. However, these compounds must be evaluated as part of the health-sessment approach (VPH/EPH).				a .
ACTION	l: Qua	alify reported TIC results as estimated and flag (NJ).				
3.:	5	If dilutions were required, were dilution factors reported?	Yes 🔼	No []	N/A []	Comments:
ACTION:	If no,	contact the lab for submission.				
4.0 <u>M</u>	etho	d Blanks				
4. ACTIO		Is the Method Blank Summary present? no, call the laboratory for submission of missing data.	Yes [V]	No [_]	N/A []	Comments:
4.	2	For the analysis of SVOCs, has a method blank been analyzed for each analysis batch of field samples of 20 or less?	Yes [V]	No [_]	N/A []	Comments:
ACTIO justifica		If no, document discrepancy in case narrative and contact lab for Consult senior chemist for action needed.				

	4.3 Is the method blank less than the PQL? Yes No NA Comments:
	E: MADEP allows common laboratory contaminants (such as phthalates) to be nt at concentrations < 5x the PQL
	4.4 Do any method blanks have positive results for SVOC parameters? Qualify data Yes No [_] N/A [_] Comments:
	For the common contaminants (phthalates):
	If the sample concentration is $< 10 \times$ blank value, flag sample result non-detect "U" at the PQL or the concentration reported if greater than the PQL. Bis(2-ethylhexylphthafafe © 0.699 mg/L is the sample concentration is $> 10 \times$ blank value, no qualification is needed. Bis(2-ethylhexylphthafafe © 0.699 mg/L is the sample concentration is $> 10 \times$ blank value, no qualification is needed.
	If the sample concentration is > 10 × blank value, no qualification is needed. Method blank × 10 = 6.9 mg/k A
	For other SVOC contaminants:
	If the sample concentration is $< 5 \times$ blank value, flag sample result non-detect "U" at the PQL or the concentration reported if greater than the PQL.
	If the sample concentration is $> 5 \times$ blank value, no qualification is needed.
	ON: For any blank with positive results, list all contaminants for each method blank, including the concentration detected and the flagging level (flagging 5x or 10x the blank value) and the associated samples and qualifiers.
6.0	Laboratory Control Standard
	5.1 Was a laboratory control standard run with each analytical batch of 20 Yes No No NA Comments: samples or less?
	ION: Call laboratory for LCS form submittal. If data are not available, use ssional judgment to determine the usability of sample results associated with that

5.2	Is a LCS Summary Form present?	Yes 🔽	No [_]	N/A []	Comments:	
ACTION:	If no, contact lab for resubmission of missing data.					
5.3	Is the recovery of any analyte outside of control limits?	Vec []	No L	N/A [1	Comments:	
NOTE: A <u>fu</u>	II target, second source LCS is required by MADEP.	165	140 [0]	IVA [comments.	
	DEP guidelines list LCS recovery limits as 40-140 for base-neutral compounds e acid compounds. The laboratory must identify analytes that routinely exceed th					a
	ecovery is above the upper limit, qualify all positive sample results within the batth as (J). If LCS recovery is <10%, non-detect results are rejected (R		overy is bel	ow the lower	limit but > 10%, q	ualify all positive and no-
	Are 80% of LCS recoveries within laboratory control limits? If 80% of LCS recoveries are not within limits, use professional judgm Senior Chemist.	Yes [<u>/</u>]	No [_]	N/A [_]	Comments:	
6.0 <u>Matr</u>	ix Spikes					
· · ·	may be collected at different frequencies based on monthly, quarterly chedules. Confirm spike requirements for each set with the senior chemi					
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6.1 were	Were project specified MS/MSDs collected? List project samples that spiked.	Yes []	No 🚺	N/A []	Comments:
ACTION:	f no, contact senior chemist to see if any were specified.				
6.2 ACTION	Is the MS/MSD recovery form present? If no, contact lab for resubmission of missing data.	Yes []	No []	N/A 🗹	Comments:
6.3	Were matrix spikes analyzed at the required frequency of 1 per 20 samples per matrix?	Yes []	No []	N/A [V]	Comments:
ACTION	If any matrix spike data are missing, call lab for resubmission.	KC \$			

6.4 Are any SVOC spike recoveries outside of the QC limits?

NOTE: %R = (SSR-SR) x 100% Where: SSR = Spiked sample result SR = Sample result SA = Spike added

SA Spike added

NOTE: A full target, second source MS/MSD is required by MADEP.

NOTE: MADEP guidelines list MS/MSD recovery limits as 40-140 for base-neutral compounds and 30-130 for acid compounds.

NOTES: 1) If only one of the recoveries for an MS/MSD pair is outside of the control limits, no qualification is necessary. Use professional judgment for the MS/MSD flags.

2) If the MS/MSD was performed by the laboratory on a non-project sample, no qualification is required.

ACTION: MS/MSD flags only apply to the sample spiked. If the recoveries of the MS and MSD exceed the upper control limit, qualify positive results as estimated (J). If the recoveries of the MS and MSD are lower than the lower control limit, qualify both positive results and non-detects (J). If LCS recovery is <10%, non-detect results are rejected (R).

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6.5 Are any RPDs for MS/MSD recoveries outside of the QC limits?	Yes No No N/A Comments:
NOTE: RPD = S-D $x = 100\%$ Where: S = MS sample result D = MSD sample result	
NOTE: MADEP guidelines list MS/MSD RPD limits for water as ≤ 20 and soils as ≤ 30.	
NOTE: Laboratory control limits apply when spiked sample results fall within the normal calibration range. If dilutions are required due to high sample concentrations, the data are evaluated, but no flags are applied.	
ACTION: If the RPD exceeds the control limit, qualify positive results and non-detects (J).	
.0 <u>Surrogate Recoveries</u>	
Were one or more SVOC surrogate recoveries outside of laboratory limits for any sample or method blank? If yes, were samples re-analyzed?	Yes No
NOTE : $%R = QD \times 100\%$ Where: $S = MS$ sample result $D = MSD$ sample result	only (1) surrogated out low in OC-GW-16
NOTE: MADEP guidelines list surrogate limits for soils as 30-130% for all surrogates, and for water as 30-130% for base-neutrals and 15-110% for acid surrogates.	Phenol-d5 13%
NOTE : Qualify BNE results based upon BNE surrogates and AE results based upon AE surrogates.	Phenol-ds 13% No Qualification necessary
ACTION: If recoveries are >10%, but 2 or more from any one fraction (acid or base-neutral) fail to meet QC criteria: (1) For recoveries below the QC limit, qualify non-detects and positives (J), and (2) For recoveries above the QC limit, qualify only positives (J). If any surrogate recovery is <10% (unless the lab QC limits are below	

10% in which case, results are flagged as stated above), flag positives (J) and reject

	letects (R).				
8.0	Sampling Accuracy				
	ajority of ground water samples are collected directly from a tap, process stream, or edicated tubing. Rinse blanks will not be collected.				
	8.1 Were rinsate blanks collected? Prior to evaluating rinsate blanks, obtain a list of the associated samples from the project chemist.	Yes []	No [V]	N/A [_]	Comments:
NOT	E: MADEP does not specify the collection of rinsate blanks.			,	
	8.2 Do any rinsate blanks have positive results?	Yes []	No []	N/A [V]	Comments:
NOT indic	E: For the common contaminants (phthalates), qualification is applied as ated above using a 10x blank value in lieu of a 5x blank value.				
	If the sample concentration is $< 5 \times$ blank value, flag sample result non-detect "U" at the PQL or the concentration reported if greater than the PQL.				
	If the sample concentration is $> 5 \times$ blank value, no qualification is needed.				
9.0	Field Duplicates				
	9.1 Were field duplicate samples collected? Obtain a list of the samples and their associated field duplicates.	Yes	No 🔼	N/A [_]	Comments:
	9.2 Were field duplicates collected per the required frequency?	Yes []	No []	N/A 🗹	Comments:
□ SO	W □ QAPP □ MADEP Option 1(1 per 20) □ MADEP Option 3 (1 per 10)				
	9.3 Was the RPD $\leq 50\%$ for soils or waters? Calculate the RPD for all results and	Yes []	No []	N/A	Comments:

	attach to this review.			
ACTIO	TION: RPD must be ≤50% for soil and water. Qualify data (J) for both sample results if the RPD exceeds 50%.			
10.0	Application of Validation Qualifiers			
	Was any of the data qualified? Yes \[No \left[] N/A \left[] Comment	ts:		
	If so, apply data qualifiers directly to the DQE copy of laboratory report and flag pages for entry in database. Ow eve defection of the database Over eve eve	of due	bis (2-ethyl hexyl) Phito blauk contamina
REFE	FERENCES			
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